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# Canine Parvovirus (CPV) ELISA

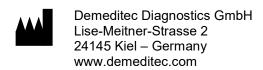




**DEPARVT0370** 



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# Demeditec Canine Parvovirus (CPV) ELISA DEPARVT0370

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#### 1. INTRODUCTION

The Canine Parvovirus (CPV) infection is a highly contagious viral illness that affects dogs. Parvoviruses are single-stranded DNA viruses with a genome of about 5000 nucleotides. They are one of the smallest known viruses. Without envelope they have a diameter of about 20-25 nm. They show a high physical stability. The capsid is highly resistant and the virions can remain infectious for months to years. The virus manifests itself in two different forms. The more common form is the intestinal form, which is characterized by vomiting, diarrhea, weight loss, and lack of appetite (anorexia). The less common form is the cardiac form, which attacks the heart muscles of very young puppies, often leading to death. Parvovirus is excreted in the feces. The most common mode of transmission is indirectly. The viruses are taken up oronasal from the contaminated environment or contaminated litter. They are transported via the lymphatic cells of the pharynx in almost all organs. Replication occurs preferably in lymphocytic cells. During replication, the host cell is destroyed. Via the epithelial cells the virus is transported in the intestinal lumen and is then excreted within the feces. The majority of cases are seen in puppies that are between six weeks and six months old.

Species	Disease	Symptoms (e.g.)	Transmission route
Canine Parvovirus (CPV)	Parvovirus in- fection of dogs	<ul> <li>Severe, bloody diarrhea</li> <li>Lethargy</li> <li>Anorexia</li> <li>Fever</li> <li>Vomiting</li> <li>Severe weight loss</li> </ul>	Oronasal

Infections may be diagnosed by:

- Cell culture
- PCR
- Serology: Determination of specific antibodies by ELISA

#### 2. INTENDED USE

The Demeditec Canine Parvovirus (CPV) ELISA is intended for the qualitative determination of antibodies against Canine Parvovirus (CPV) in veterinary serum.

#### 3. PRINCIPLE OF THE ASSAY

The qualitative immunoenzymatic determination of specific antibodies is based on the ELISA (Enzyme-linked Immunosorbent Assay) technique. Microtiterplates are coated with specific antigens to bind corresponding antibodies of the sample. After washing the wells to remove all unbound sample material a horseradish peroxidase (HRP) labelled conjugate is added. This conjugate binds to the captured antibodies. In a second washing step unbound conjugate is removed. The immune complex formed by the bound conjugate is visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product. The intensity of this product is proportional to the amount of specific antibodies in the sample. Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450/620 nm is read using an ELISA Microtiterplate reader.

#### 4. MATERIALS

# 4.1. Reagents supplied

- 1. **SORB** MT Microtiterplate: 12 break-apart 8-well snap-off strips coated with Canine Parvovirus (CPV) antigens; in resealable aluminium foil.
- 2. SAM DIL Sample Dilution Buffer: 1 bottle containing 100 mL of phosphate buffer (10 mM) for sample dilution; pH 7.2 ± 0.2; coloured yellow; ready to use; white cap; ≤ 0.0015 % (v/v) CMIT/MIT (3:1).
- 3. STOP SOLN Stop Solution: 1 bottle containing 15 mL sulphuric acid, 0.2 mol/L; ready to use; red cap.
- 4. **WASH SOLN 20x Washing Buffer (20x conc.):** 1 bottle containing 50 mL of a 20-fold concentrated phosphate buffer (0.2 M), pH 7.2 ± 0.2, for washing the wells; white cap.
- 5. **ENZ CONJ Conjugate:** 1 bottle containing 20 mL of peroxidase labelled Protein A/G; coloured yellow; ready to use; white cap; ≤ 0.02 % (v/v) MIT.
- 6. **SUB TMB TMB Substrate Solution:** 1 bottle containing 15 mL 3,3',5,5'-tetramethylbenzidine (TMB), < 0.1 %; ready to use; yellow cap.
- CAL C Positive Control: 1 vial containing 2 mL; coloured yellow; ready to use; red cap; ≤ 0.02 % (v/v) MIT.
- 8. CAL B Cut-off Control: 1 vial containing 3 mL; coloured yellow; ready to use; green cap; ≤ 0.02 % (v/v) MIT.
- 9. CAL A Negative Control: 1 vial containing 2 mL; coloured yellow; ready to use; blue cap; ≤ 0.0015 % (v/v) CMIT/MIT (3:1).

For hazard and precautionary statements see 12.1

For potential hazardous substances please check the safety data sheet.

#### 4.2. Materials supplied

- 1 Cover foil
- 1 Instruction for use (IFU)

#### 4.3. Materials and Equipment needed

- ELISA Microtiterplate reader, equipped for the measurement of absorbance at 450/620 nm
- Incubator 37 °C
- Manual or automatic equipment for rinsing Microtiterplate wells
- Pipettes to deliver volumes between 10 and 1000 μL
- Vortex tube mixer
- Distilled water
- Disposable tubes

#### 5. STABILITY AND STORAGE

Store the kit at 2...8 °C. The opened reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

#### 6. REAGENT PREPARATION

It is very important to bring all reagents and samples to room temperature (20...25 °C) and mix them before starting the test run!

#### 6.1. Microtiterplate

The break-apart snap-off strips are coated with Canine Parvovirus (CPV) antigens. Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8 °C.

#### 6.2. Washing Buffer (20x conc.)

Dilute Washing Buffer 1 + 19; e. g. 10 mL Washing Buffer + 190 mL distilled water. The diluted buffer is stable for 5 days at room temperature (20...25 °C). In case crystals appear in the concentrate, warm up the solution to 37 °C e.g. in a water bath. Mix well before dilution.

#### 6.3. TMB Substrate Solution

The reagent is ready to use and has to be stored at 2...8 °C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away.

#### 7. SAMPLE COLLECTION AND PREPARATION

Use canine serum samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

#### 7.1. Sample Dilution

Before assaying, all samples should be diluted 1+100 with Sample Dilution Buffer. Dispense 10  $\mu$ L sample and 1 mL Sample Dilution Buffer into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

#### 8. ASSAY PROCEDURE

Please read the instruction for use carefully **before** performing the assay. Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend increasing the washing steps from three up to five and the volume of Washing Buffer from 300  $\mu$ L to 350  $\mu$ L to avoid washing effects. Pay attention to chapter 12. Prior to commencing the assay, the distribution and identification plan for all samples and standards/controls (duplicates recommended) should be carefully established on the plate layout. Select the required number of microtiter strips or wells and insert them into the holder.

Perform all assay steps in the order given and without any delays.

A clean, disposable tip should be used for dispensing each standard/control and sample. Adjust the incubator to  $37 \pm 1$  °C.

- 1. Dispense 100 µL standards/controls and diluted samples into their respective wells. Leave well A1 for the Substrate Blank.
- 2. Cover wells with the foil supplied in the kit.
- 3. Incubate for 1 hour ± 5 min at 37 ± 1 °C.
- 4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300  $\mu$ L of Washing Buffer. Avoid overflows from the reaction wells. The interval between washing and aspiration should be > 5 sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!
  - Note: Washing is important! Insufficient washing results in poor precision and false results.
- 5. Dispense 100  $\mu$ L Conjugate into all wells except for the Substrate Blank well A1.
- 6. Incubate for 30 min at room temperature (20...25 °C). Do not expose to direct sunlight.
- 7. Repeat step 4.
- 8. Dispense 100 µL TMB Substrate Solution into all wells.
- 9. **Incubate for exactly 15 min at room temperature (20...25 °C) in the dark.** A blue colour occurs due to an enzymatic reaction.
- 10. Dispense 100 µL Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution, thereby a colour change from blue to yellow occurs.
- 11. Measure the absorbance at 450/620 nm within 30 min after addition of the Stop Solution.

#### 8.1. Measurement

Adjust the ELISA Microtiterplate reader to zero using the Substrate Blank.

If - due to technical reasons - the ELISA Microtiterplate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results!

**Measure the absorbance** of all wells at **450 nm** and record the absorbance values for each standard/control and sample in the-plate layout.

Bichromatic measurement using a reference wavelength of 620 nm is recommended.

Where applicable calculate the **mean absorbance values** of all duplicates.

#### 9. RESULTS

#### 9.1. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

- Substrate Blank: Absorbance value < 0.100
- Negative Control: Absorbance value < 0.200 and < Cut-off</li>
- Cut-off Control: Absorbance value 0.150 1.300
- Positive Control: Absorbance value > Cut-off

If these criteria are not met, the test is not valid and must be repeated.

#### 9.2. Calculation of Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations.

Example: Absorbance value Cut-off Control 0.44 + absorbance value Cut-off control 0.42 = 0.86 / 2 = 0.43

Cut-off = 0.43

# 9.2.1. Results in Units [U]

Sample (mean) absorbance value x 10 = [Units = U]

Cut-off

Example:  $1.591 \times 10 = 37 \text{ U}$ 

0.43

#### 9.3. Interpretation of Results

Normal value ranges for this ELISA should be established by each laboratory based on its own sample populations in the geographical areas serviced. The following values should be considered as a guide-line:

Cut-off	10 U	-
		Antibodies against the pathogen are present. There has been a contact with the antigen (pathogen resp. vaccine).
Equivocal 9 – 11 U		Antibodies against the pathogen could not be detected clearly. It is recommended to repeat the test with a fresh sample in 2 to 4 weeks. If the result is equivocal again the sample is judged as <b>negative</b> .
Negative	< 9 U	The sample contains no antibodies against the pathogen. A previous contact with the antigen (pathogen resp. vaccine) is unlikely.

#### 10. SPECIFIC PERFORMANCE CHARACTERISTICS

The results refer to the groups of samples investigated; these are not guaranteed specifications. The performance data have been established with canine samples. Due to the nature of the Protein A/G conjugate this ELISA should react with other mammalian species also. More detailed information is available on request.

#### 10.1. Precision

<u>Intraassay</u>	n	Mean (E)	CV
(%)			
#1	24	0.350	8.96
#2	24	1.211	2.60
#3	24	0.777	8.61

<u>Interassay</u>	n	Mean (U)	CV (%)
#1	12	4.53	8.34
#2	12	23.00	4.63
#3	12	15.24	7.78

#### 10.2. Specificity

The specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte.

Specificity canine: > 98 % (95 % confidence interval: 54.07 % - 100 %)

#### 10.3. Sensitivity

The sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte.

Sensitivity canine: 98.21 % (95 % confidence interval: 90.45 % - 99,95 %)

#### 10.4. Interferences

Interferences with hemolytic, lipemic or icteric samples are not observed up to a concentration of 10 mg/mL hemoglobin, 5 mg/mL triglycerides and 0.5 mg/mL bilirubin.

# 10.5. Cross Reactivity

Cross reactions cannot be excluded.

## 11. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or repeated freeze-thaw cycles of the sample may affect the absorbance values.

#### 12. PRECAUTIONS AND WARNINGS

- Only for research use.
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- All components of human origin used for the production of these reagents have been tested for <u>anti-HIV</u> antibodies, anti-HCV antibodies and HBsAg and have been found to be non-reactive.
- Do not interchange reagents or strips of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette patient samples and dispense reagents without splashing accurately into the wells.
- The ELISA is only designed for qualified personnel who are familiar with good laboratory practice.

#### 12.1. Safety note for reagents containing hazardous substances

Reagents may contain CMIT/MIT (3:1) or MIT (refer to 4.1)

Therefore, the following hazard and precautionary statements apply.

warning
$\langle \cdot \rangle$

	may cause an anergie chin reaction.
P261	Avoid breathing spray
P280	Wear protective gloves/protective clothing.
P302+P352	IF ON SKIN: Wash with plenty of soap and water.

May cause an allergic skin reaction

P333+P313 If skin irritation or rash occurs: Get medical advice/attention.

P362+P364 Take off contaminated and Wash it before reuse.

Further information can be found in the safety data sheet.

#### 12.2. Disposal Considerations

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Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

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#### **ABBREVIATIONS**

CMIT	5-chloro-2-methyl-4-isothiazolin-3-one
MIT	2-methyl-2H-isothiazol-3-one

## **SUMMARY OF TEST PROCEDURE**

# **SCHEME OF THE ASSAY**

Canine Parvovirus (CPV) ELISA

# **Test Preparation**

Prepare reagents and samples as described.

Establish the distribution and identification plan for all samples and standards/controls.

Select the required number of microtiter strips or wells and insert them into the holder.

**Assay Procedure** 

Assay Procedure						
	Substrate Blank (A1)	Negative Control	Cut-off Control	Positive Control	Sample (diluted 1+100)	
Negative Control	-	100 µL	-	-	-	
Cut-off Control	-	-	100 μL	-	-	
Positive Control	-	-	-	100 µL	-	
Sample (diluted 1+100)	-	-	-	-	100 μL	
Cover wells with foil supplied in the kit  Incubate for 1 h at 37±1 °C  Wash each well three times with 300 µL of Washing Buffer						
Conjugate	-	100 µL	100 μL	100 μL	100 µL	
Incubate for 30 min at room temperature (2025 °C)  Do not expose to direct sunlight  Wash each well three times with 300 µL of Washing Buffer						
TMB Substrate So- lution	100 μL	100 µL	100 μL	100 μL	100 μL	
Incubate for exactly 15 min at room temperature (2025 °C) in the dark						
Stop Solution	100 μL	100 μL	100 μL	100 μL	100 μL	
Photometric measurement at 450 nm (reference wavelength: 620 nm)						

# SYMBOLS USED WITH DEMEDITEC ASSAYS

Symbol	English	Deutsch	Française	Espanol	Italiano
CE	European Conformity	CE-Konformitäts- kennzeichnung	Conforme aux normes européennes	Conformidad europea	Conformità europea
Ţ <u>i</u>	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instruc- tions d'utilisation	Consulte las Instrucciones	Consultare le istruzioni per l'uso
IVD	In vitro diagnostic de- vice	In-vitro-Diagnostikum	utilisation Diagnostic in vitro	Diagnóstico in vitro	Per uso Diagnostica in vitro
RUO	For research use only	Nur für Forschungs- zwecke	Seulement dans le cadre de recherches	Sólo para uso en investigación	Solo a scopo di ricerca
REF	Catalogue number	Katalog-Nr.	Référence	Número de catálogo	No. di catalogo
LOT	Lot. No. / Batch code	Chargen-Nr.	No. de lot	Número de lote	Lotto no
$\Sigma$	Contains sufficient for <n> tests/</n>	Ausreichend für "n" Ansätze	Contenu suffisant pour "n" tests	Contenido suficiente para <n> ensayos</n>	Contenuto sufficiente per "n" saggi
$\triangle$	Note warnings and pre- cautions	Warnhinweise und Vorsichtsmaßnahmen beachten	Avertissements et me- sures de précaution font attention	Tiene en cuenta advertencias y precauciones	Annoti avvisi e le pre- cauzioni
	Storage Temperature	Lagerungstemperatur	Température de con- servation	Temperatura de conservacion	Temperatura di conservazione
$\square$	Expiration Date	Mindesthaltbarkeits- datum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
***	Legal Manufacturer	Hersteller	Fabricant	Fabricante	Fabbricante
Distributed by	Distributed by	Vertrieb durch	Distribution par	Distribución por	Distribuzione da parte di
V <x></x>	Version	Version	Version	Versión	Versione
2	Single-use	Einmalverwendung	À usage unique	Uso único	Uso una volta